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GVS

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Evaluation of the impact of birth litter size on birth weight and subsequent birth-to-market performance

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Table 1: Effect of birth litter size on piglets characteristics at birth, least squares treatment means

Birth litter size class	1-9	10-12	13-14	15-16	17&Up	SEM*
Average parity	2.85[a]	2.70[a]	2.81[a]	2.88[a]	3.12[b]	0.070
Mean piglet birth weight (lbs)	3.62[ce]	3.34[d]	3.11[c]	2.96[b]	2.74[a]	0.013
Within litter variation†	0.434[a]	0.535[b]	0.606[b]	0.635[c]	0.650[c]	0.0099
Prewean mortality (mortality as % of total born alive)††	9.01	10.19	14.55	16.32	21.63	
Nursery ADG (lbs/day)	1.25[d]	1.23[c]	1.21[b]	1.21[b]	1.18[a]	0.004
Nursery mortality (mortality as % of pigs placed)††	1.32	1.47	1.04	1.15	1.71	
Finish ADG (lbs/day)	2.20[c]	2.16[ab]	2.17[b]	2.15[a]	2.15[a]	0.010
Finisher mortality (mortality as % of pigs placed)††	1.27	1.70	2.24	2.05	1.90	

[a,b,c,d,e] Values with different superscripts within row are significantly different ($P < .05$)

* Maximum value

† Standard deviation of individual pig birth weights within birth litter

†† Raw means

Association of gilt vulval score with follicle development and expression of estrus

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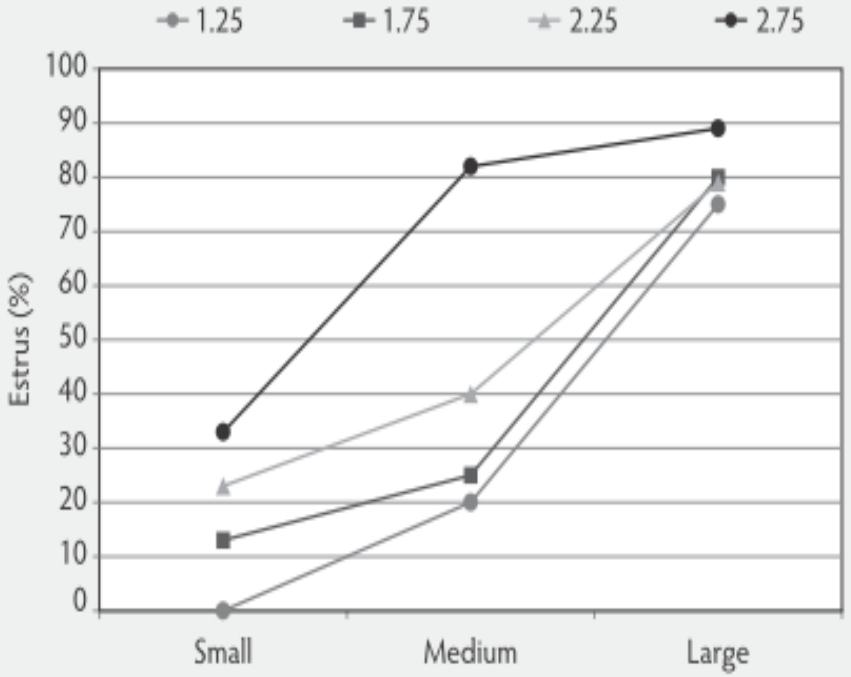
Table 1: Percentage of gilts with average of the largest follicle sizes in relation to average vulval score during the follicle phase.

	Small	Medium	Large
1.25	0%	20%	75%
1.75	13%	25%	80%
2.25	23%	40%	79%
2.75	33%	82%	89%

Average vulva score over days (0, 1, 2 or 3) of the follicle phase were classified as no swelling (0), minimal swelling (1), moderate swelling (2), or considerable swelling(3)). The ranges were the midpoints 1.0 to 1.5, >1.49 to 1.99, 2.0 to 2.49 and ≥2.5), highest vulva score during observation, and average size of the largest follicles (small (< 3 mm), medium (3 to 6.49 mm) and large (> 6.49 mm)).

Figure 1: Average vulval score during the follicle phase in relation to average size of the largest follicles and its relation to expression of estrus in replacement gilts.



Timed insemination following intravaginal OvuGel® treatment in postpartum sows

Paul Yeske[1], DVM; Michael Johnston[2], MS; Mark Swanson[2], PhD;
Stephen Webel[2], PhD; Robert Kraeling[3], PhD
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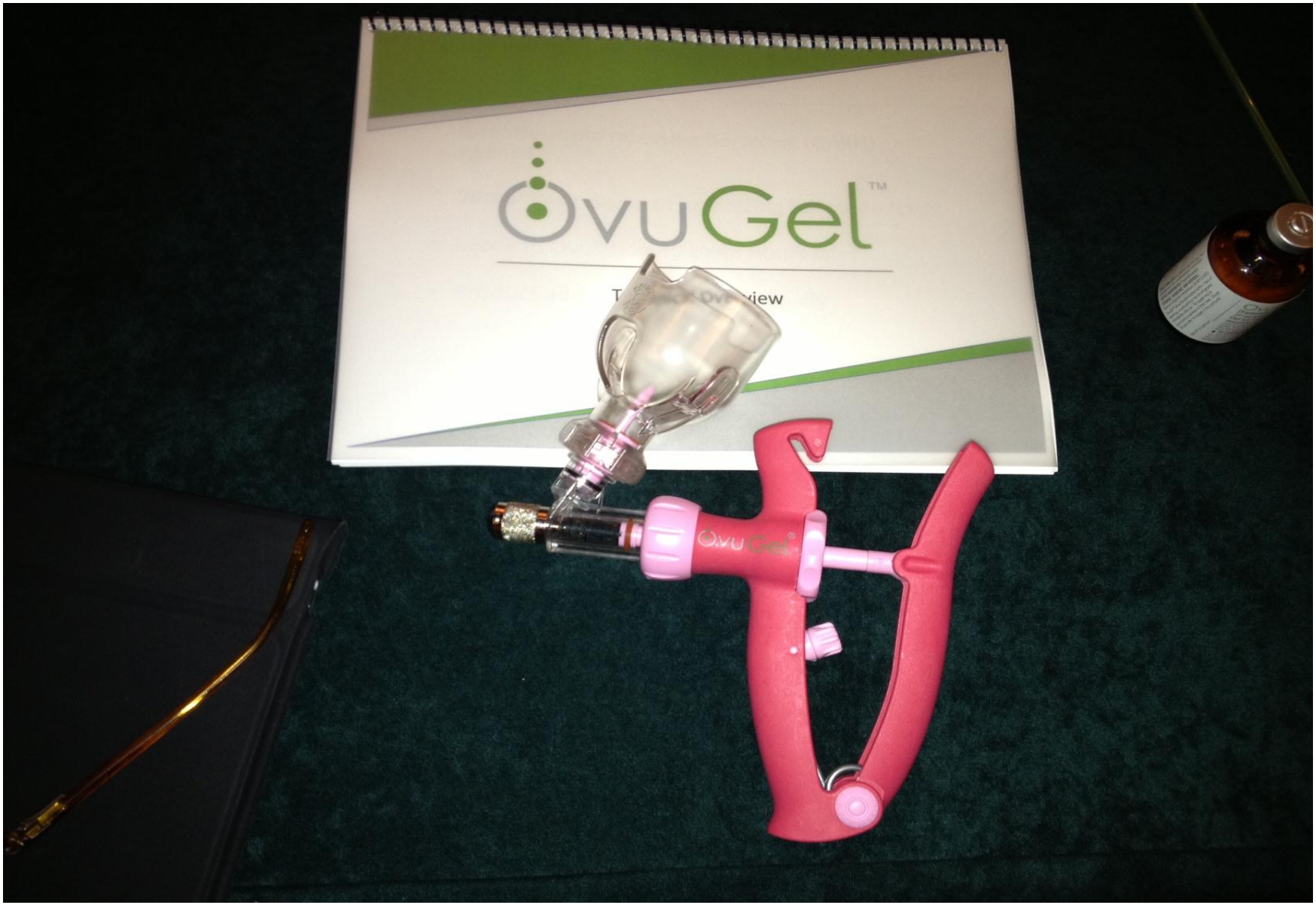
[3]L&R Research Associates, Inc., Watkinsville, Georgia

Table 1: Responses to treatment (LS means ± SEM) and comparisons between Control and OvuGel® sows[1].

	Control	OvuGel®	SEM	Contrast, *P* <
Sows allocated to treatments	198	197	*	*
No. of sows inseminated within 7-d after weaning	181	197	*	*
No. of inseminations/sow	2.2	1.0	0.02	0.0001
Percentage of allotted sows pregnant at 30 days	82.3	84.3	2.67	0.61
No. sows farrowed	154	157	*	*
Percentage of allotted sows farrowed	78.2	79.7	2.92	0.74
No. of pigs born/semen dose	5.2	9.7	0.34	0.0001
No. of pigs born/litter	12.7	12.2	0.25	0.22
No. of pigs born alive/litter	11.8	11.3	0.24	0.17

[1] Control sows bred following normal farm SOP. OvuGel® sows were treated between 97 and 102 hours post-weaning and inseminated once 21-22 hours later. LS means calculated using the PROC MIX procedure of SAS and assumed a model with treatment, replication, and treatment by replication interactions.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Economic value of feed efficiency and body condition scoring in a sow herd

John Sonderman, PhD; Thomas Rathje, PhD
Danbred North America, Columbus, Nebraska

Gilt management 2.0

Juan Carlos Pinilla, DVM, MS; Rodrigo Teuber, DVM; Jose Piva, DVM; James Coates, BSc
PIC Technical Services - Hendersonville, Tennessee

Group sow performance; comparing three group housed systems to conventional stalls

Jeff Schoening
Automated Production System

- Scrofe in box: 0,227 kg giorno in più di consumo mangime
- Scrofe con auto alimentatore: 0,114 Kg di mangime in meno al giorno
- Mediamente da gabbia a box il costo di mangime del lattone aumenta di 0,57 $.
- La produttività risulta sovrapponibile nei due metodi

Quantifying the production impact in farms going through load-close-expose programs for PRRS virus

Daniel Linhares; Montserrat Torremorell; Robert Morrison
SDEC, CVM, University of Minnesota, Saint Paul, Minnesota

Time to produce PRRSv-negative pigs from infected breeding sites

Daniel Linhares; Montserrat Torremorell; Robert Morrison
SDEC, CVM, University of Minnesota, Saint Paul, Minnesota

PRRS virus elimination: How to prove herd negativity?

Montserrat Torremorell, DVM, PhD; Bob Morrison, DVM, PhD
College of Veterinary Medicine, University Of Minnesota, St. Paul, Minnesota

Negatività per PRRS dopo la chiusura di un allevamento

- In queste situazioni le scrofette più giovani sono gli animali più suscettibili; allo stesso modo, per eseguire verifiche sierologiche, sono indicati i suinetti da svezzare
- Se ci aspettiamo una prevalenza bassa (< del 10%) dobbiamo aumentare fino a 60 campioni (5% prevalenza).
-Dopo la reintroduzione degli animali negativi, questi rappresentano una popolazione ideale da testare. A partire da 30 gg dopo l'ingresso
- Frequenza: mensile e durata almeno di 4 mesi

Evaluation of biosecurity measures and management variables as risk factors for infection of growing pigs that are negative at placement with porcine reproductive and respiratory syndrome virus

Caleb Robb; Derald Holtkamp, DVM, MS; Paul Yeske, DVM, MS; Aaron Lower, DVM;
James Lowe, DVM, MS; Dale Polson, DVM, MS; Peter Lasley, DVM, MS

- Accurato lavaggio e disinfezione del capannone
- Trasporti: lavaggio e asciugatura del camion. Proprietà o meno del camion.
- Frequenza di visita all'allevamento
- Procedure igieniche per personale che visita l'allevamento
- Ore di "astinenza" da altri suini
- Densità suina della zona

PRRSv half-life in manure

Daniel Linhares, DVM, MBA, PhD; Han Soo Joo, DVM, PhD;
Montserrat Torremorell, DVM, PhD; Robert Morrison, DVM, MBA, PhD
Swine Disease Eradication Center, College of Veterinary Medicine,
University of Minnesota, Saint Paul, Minnesota

Resistenza del virus PRRS nel liquame, espresso come emivita

- 113 ore a 4°C; 15 ore a 20°C; 1,6 ore a 40°C; 2,9 minuti a 60°C; 0,36 minuti a 80°C.

Nelle sperimentazioni in campo i materiali contaminati di liquame sono stati mantenuti in ambiente chiuso in modo da conservare l'umidità, fattore favorevole alla conservazione del virus. E si è visto come le alte temperature di trattamento consentono un'asciugatura più veloce e quindi una emivita inferiore del virus.

> **Utilizzare acqua calda e/o un'asciugatura con aria calda durante la bonifica dei mezzi di trasporto !**

Field investigations of perceived PCV2 vaccine failures and experimental challenge trials that support transitioning to a PCV2b-based vaccine

Tanja Opriessnig[1], Dr med vet, PhD; Kevin O'Neill[1], MS; Priscilla Gerber[1], DVM, PhD; Luis G. Giménez-Lirola[1], PhD; Lei Zhou[2], PhD; Nathan Beach[2], PhD; X.J. Meng[2], MD, PhD; Patrick Halbur[1], DVM, MS, PhD

[1]Department of Veterinary Diagnostic and Production Animal Medicine, College of Veterinary Medicine, Iowa State University, Ames, Iowa; [2]Department of Biomedical Sciences and Pathobiology, Virginia Polytechnic Institute and State University, Blacksburg, Virginia

The influence of fetal PCV2 infection response on lifetime performance and vaccine efficacy

James F. Lowe[1,2], DVM, MS; Laura Greiner[3], PhD

[1]Lowe Consulting, Ltd. , Albers Illinois; [2]Department of Veterinary Clinical Medicine, University of Illinois, Urbana, Illinois; [3]Innovative Swine Solutions LLC, Carthage, Illinois

Comparison of different *Mycoplasma hyopneumoniae* vaccination schedules in growing pigs with a seeder pig challenge

Angie Supple[1], DVM; Amber Stricker[1], DVM, MS; Amanda Ness Sponheim[2], DVM
[1]Suidae Health and Production, Algona, Iowa;
[2]Boehringer Ingelheim Vetmedica, Inc., St. Joseph, Missouri

Comparative efficacy of Ingelvac MycoFLEX® vaccination in pigs at 3 weeks of age from Ingelvac MycoFLEX® vaccinated and non-vaccinated sows on the reduction of lung lesions

Jose Angulo, DVM; Greg Cline, DVM; Kari Saddoris-Clemons, PhD
Boehringer Ingelheim Vetmedica, Inc.

PRRSv control in finisher pigs, a large scale barn study in a high dense area in USA

Rebecca Robbins[1], DVM; Perry Harms[2], DVM, MS; Jose Angulo[3], DVM; Alan Scheidt[3], DVM, MS; Reid Philips[3], DVM; John Kolb[3], DVM, Dipl ABVP
[1]Murphy-Brown LLC Eastern Operations; [2]Smithfield Premium Genetics Group;
[3]Boehringer Ingelheim Vetmedica Inc.

INFLUENZA

Alcuni messaggi dalle relazioni:

•Il virus può passare da animali positivi a negativi con fomiti, al di l di qualsiasi livello di biosicurezza adottata

•La circolazione del virus della mandria infetta è molto dinamica: scrofa > suinetto > scrofa. Le scrofette di rimonta possono sia introdurre il virus sia agire da “luogo di replicazione” di un virus residente e a loro sconosciuto.

•La vaccinazione, nella situazione americana, è un mezzo per la riduzione dei sintomi clinici; può dare garanzie solo parziali. Oggi stanno provando vaccini autogeni da isolamenti recenti.

•Quando si usa nei siti 1, meglio a tappeto che applicata prima del parto

•Nei siti 3, visto la efficacia parziale dei vaccini, puntano sulla qualità dell’ambiente e sulla qualità del personale: la terapia individuale è quella che consente di contenere i danni economici.

•Diagnosi: Fluidi orali (corde di cotone nel box) = tampone nasale

….. e non ho altro da dire su questa faccenda.

(Forrest Gump)

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